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- molecules to disease, R. C. A. Thompson, J. A. Reynoldson, and A. J. Lymbery (eds.). CAB International, Wallingford, U.K., p. 15–37.
- FAYER, R., J. M. TROUT, E. WALSH, AND R. COLE. 2000. Rotifers ingest oocysts of *Cryptosporidium parvum*. *Journal of Eukaryotic Microbiology* **47**: 161–163.
- GILBERT, J. J., AND P. L. STARKWEATHER. 1977. Feeding in the rotifer *Brachionus calyciflorus*. I. Regulatory mechanisms. *Oecologia* **28**: 125–131.
- , AND ———. 1978. Feeding in the rotifer *Brachionus calyciflorus*. III. Direct observations on the effects of food type, food density, and starvation on the incidence of pseudotracheal screening. *Internationale Vereinigung fuer Theoretische und Angewandte Limnologie Verhandlungen* **20**: 2382–2388.
- KILANI, R. T., AND L. SEKLA. 1987. Purification of *Cryptosporidium* oocysts and sporozoites by cesium chloride and percoll gradients. *American Journal of Tropical Medicine and Hygiene* **36**: 505–508.
- KUHLE, K., AND W. KLEINOW. 1990. Glycosidases in *Brachionus plicatilis* (Rotifera). *Comparative Biochemistry and Physiology. B: Comparative Biochemistry* **95**: 393–402.
- NOGRADY, T., R. L. WALLACE, AND T. W. SNELL. 1993. Rotifera. Vol. 1: Biology, ecology, and systematics. *In* Guides to the identification of the microinvertebrates of the continental waters of the world, H. J. Dumont (ed.). SPB Academic Publishing, The Hague, The Netherlands, p. 1–142.
- RESVOI, P. 1926. Observations on the feeding of rotifers. *Travaux de la Societe des Naturalistes de Leningrad* **56**: 73–89. [In Russian.]
- SCHRAMM, U. 1978. Studies of the ultrastructure of the rotifer *Habrotricha rosa* Donner (Aschelminthes). The alimentary tract. *Cell and Tissue Research* **189**: 525–535.
- SEGBERS, H. 2002. Contributions to the nomenclature of Rotifera: Annotated checklist of valid family- and genus-group names. *Journal of Natural History* **36**: 631–640.
- STARKWEATHER, P. L., AND J. J. GILBERT. 1977. Radiotracer determination of feeding in *Brachionus*: The importance of gut passage times. *Archiv Fur Hydrobiologie Beiheft* **8**: 261–263.
- WALLACE, R. L., AND T. W. SNELL. 2001. Rotifera. Chapter 8. *In* Ecology and classification of North American freshwater invertebrates, 2nd ed., J. Thorp and A. Covich (eds.). Academic Press, New York, p. 195–254.

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Toxoplasmosis in Three Species of Native and Introduced Hawaiian Birds

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ABSTRACT: *Toxoplasma gondii* was found in endemic Hawaiian birds, including 2 nene geese (*Nesochen sandvicensis*), 1 red-footed booby (*Sula sula*), and an introduced bird, the Erckels francolin (*Francolinus erckelii*). All 4 birds died of disseminated toxoplasmosis; the parasite was found in sections of many organs, and the diagnosis was confirmed by immunohistochemical staining with anti-*T. gondii*-specific polyclonal antibodies. This is the first report of toxoplasmosis in these species of birds.

Toxoplasma gondii can cause mortality and subclinical infections in many species of warm-blooded animals including birds (Dubey and Beattie, 1988; Literák et al., 1992; Dubey, 2002). In tropical island ecosystems, *T. gondii* has been documented only on islands with feral cats, underlining the fact that felids are the only known definitive hosts (Wallace et al., 1972). In Hawaii, *T. gondii* has significantly affected reintroduction programs for the endangered Hawaiian crow (Work et al., 2000). This article documents acute toxoplasmosis in 3 other species of endemic and introduced Hawaiian birds, including nene goose (*Nesochen sandvicensis*), the red-footed booby (*Sula sula*), and Erckels francolin (*Francolinus erckelii*).

Birds were submitted refrigerated within 12–24 hr of death to the Hawaii Field Station or the Hawaii Department of Fish and Wildlife. Birds were weighed and examined systematically externally and internally. Tissues including brain, heart, lung, skeletal muscle, kidney, spleen, adrenal, small and large intestines, trachea, and liver were fixed in 10% buffered formalin, sectioned at 5 µm, and stained with hematoxylin and eosin for microscopic examination. Immunohistochemistry was done on paraffin-embedded sections using polyclonal anti-*T. gondii* and anti-*Sarcocystis neurona* antibodies and anti-BAG-1 antibodies specific for bradyzoites (Lindsay and Dubey, 1989; McAllister et al., 1996; Dubey and Hamir, 2000).

Two endangered Hawaiian goose (nene) goslings housed in a sand enclosure were found dead 3 days apart in August 1994 at a private zoo on Maui. Gross examination revealed both birds (1 male, 1 female) to be in excellent body condition with adequate fat reserves. The only significant gross lesion in the female included focal congestion of the

liver, marked splenomegaly, heavy wet lungs, and locally extensive fibrinous exudates on the jejunal mucosa. The main gross lesions in the male included marked hemorrhage and consolidation of the left lung, marked splenomegaly, and linear brown areas of discoloration on the large intestinal mucosa.

Histology in the female gosling revealed severe interstitial mononuclear pneumonia with intracytoplasmic tachyzoites, focal lytic hepatic necrosis, focal neuronal necrosis, severe lymphoid depletion, and severe, diffuse, nonsuppurative inflammation and necrosis of the myocardium and skeletal muscles associated with tachyzoites. There also was full-thickness necrosis of the intestinal wall extending through the lamina propria and muscularis associated with marked mononuclear response and tachyzoites. Significant microscopic findings in the male gosling included focal necrosis and marked mononuclear interstitial infiltrates associated with tachyzoites in the lung, mild focal necrosis and nonsuppurative inflammation associated with myocardial necrosis, severe lymphoid depletion, and tachyzoites in the spleen and tissue cysts with bradyzoites in the adrenal. Tissue cysts positive for BAG-1 were seen in the brain, lung, and heart (Fig. 1A–C). The brain was not examined.

In February 1995, a free-ranging Erckel's francolin was found dead on Kauai. This was a female adult in excellent body condition. Significant gross lesions included focal discoloration of the liver and heart, heavy wet mottled lungs, and a crop distended with ingesta. Histology revealed mild focal acute hepatocellular necrosis, severe myocardial necrosis and mononuclear inflammation, mild focal cerebral gliosis, and severe diffuse splenic necrosis associated with tachyzoites (Fig. 1D).

The fourth case was a red-footed booby adult that was found weak on the Oahu Island in November 1998. The bird was admitted for rehabilitation at a local zoological park and died 40 days later. Significant gross lesions included heavy wet lungs that sank in formalin, friable spleen, and engorged cerebral vasculature. Histology revealed marked diffuse necrosis and mononuclear inflammation in the lungs associated with tachyzoites, suppurative periportal inflammation of the liver, severe mononuclear inflammation associated with tachyzoites in the heart, focal neuronal necrosis of the cerebrum associated with bradyzoites,

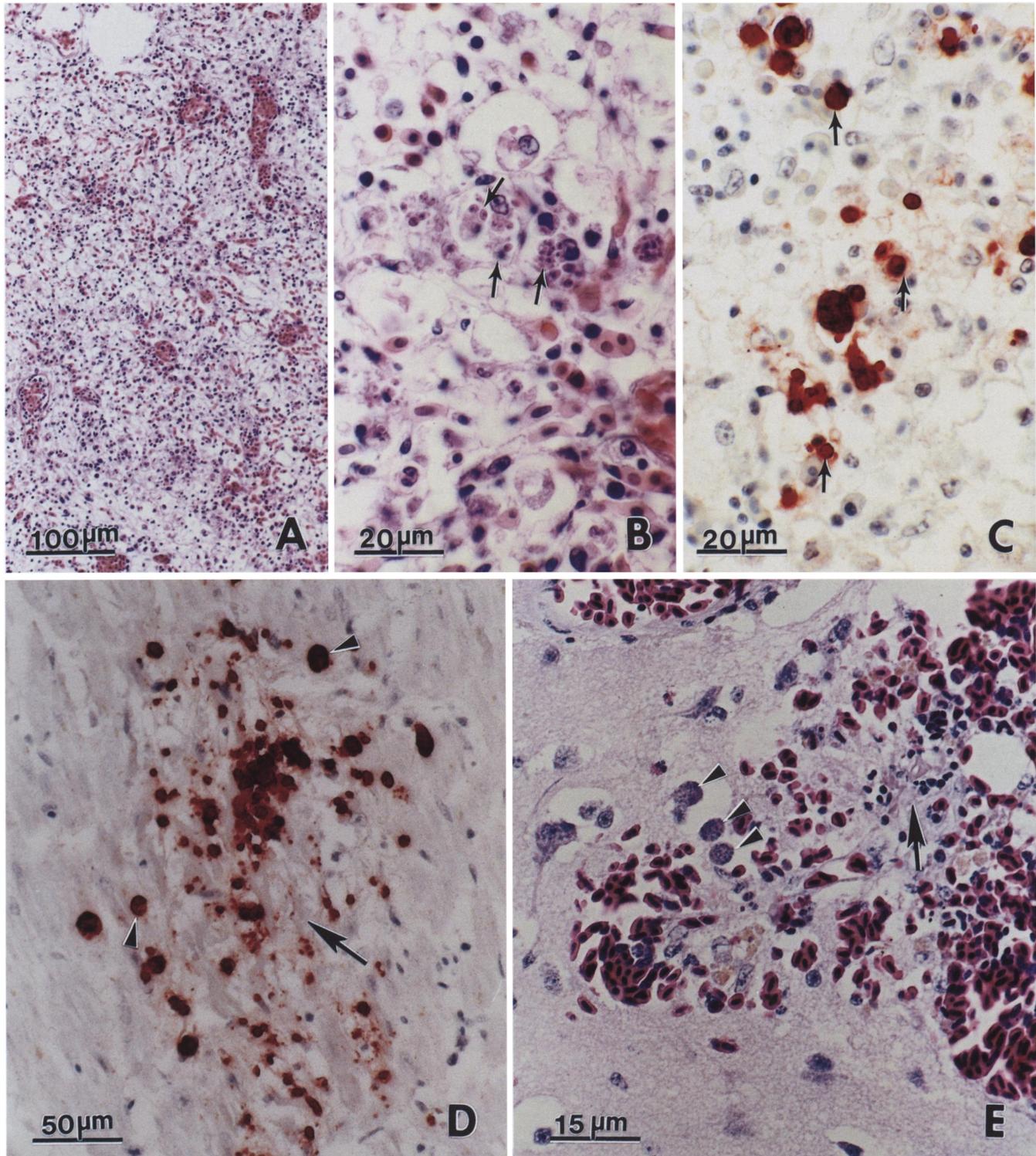


FIGURE 1. *Toxoplasma gondii* and associated lesions in birds from Hawaii. **A**, **B**, and **E**, hematoxylin and eosin stain; **C** and **D**, immunohistochemical stain with anti-*T. gondii* antibodies. (**A–C**) Lungs of nene goose. Note necrosis, infiltration by mononuclear cells, and individual tachyzoites (arrows) and groups of tachyzoites. Tachyzoites in **B** appear half the size of those in **C**. (**D**) Necrosis of myocardium of Erckel's francolin. Numerous tachyzoites (arrows) are in the lesion. (**E**) Cerebrum of red-footed booby. Note perivascularitis (arrow) and 3 tissue cysts (arrowheads).

prominent mononuclear cell hyperplasia in the spleen, and focal necrosis and nonsuppurative inflammation in the adrenal (Fig. 1E). Tissue cysts positive for BAG-1 were seen in intestines, lung, brain, and heart. Protozoans in all 4 birds reacted to anti-*T. gondii* but not to anti-*S. neurona* polyclonal antibodies.

The nene acquired infection in captivity, and it is likely that their environment (sand substrate) was conducive to the spread of toxoplasmosis by ingested feces from feral cats that were commonly observed in the vicinity. The source of infection for the francolin and the booby is less clear. The francolin may have acquired *T. gondii* through ingestion of oocysts from cat feces or a transport host (Wallace, 1973). We suspect that the booby acquired infection in the wild and not in captivity. Birds undergoing rehabilitation at the zoological park are caged and housed on concrete, which is cleaned daily, thus minimizing potential exposure to cat feces. On the other hand, feral cats are common around the largest red-footed booby colony on Oahu at the Kaneohe Marine Corps Air Station, and we suspect that the bird acquired the infection there.

Lesions in these birds were similar to those of other avian species infected with *T. gondii* (Dubey, 2002). The presence of tachyzoites in multiple organs suggested that the disease was more fulminant in the geese and francolin, and the presence of ingesta in the proventriculus of the latter suggested acute death. Presence of bradyzoites in the brain of the booby suggested a more chronic infection. In all cases, the heart and lung were the organs most commonly and severely affected.

In addition to Hawaii (Work et al., 2000), this article extends the geographic range of avian toxoplasmosis to 3 more islands (Maui, Kauai, and Oahu). Feral cats are present on all these islands, thereby providing opportunities for infection of avian hosts. This article also reinforces the fact that *T. gondii* is capable of infecting a wide variety of birds with disparate life histories, including upland game birds (Francolin), pelagic seabirds (Booby), and low-elevation-grazing anseriforms (nene geese). The fact that so few birds of these species have been documented with *T. gondii* suggests that these cases were accidental

infections. To date, we have no evidence that *T. gondii* poses a severe threat to populations of these birds.

LITERATURE CITED

- DUBEY, J. P. 2002. A review of toxoplasmosis in wild birds. *Veterinary Parasitology* **106**: 121–153.
- , AND C. P. BEATTIE. 1988. *Toxoplasmosis of animals and man*. CRC Press, Boca Raton, Florida, 220 p.
- , AND A. N. HAMIR. 2000. Immunohistochemical confirmation of *Sarcocystis neurona* infections in raccoons, mink, cat, skunk, and pony. *Journal of Parasitology* **86**: 1150–1152.
- LINDSAY, D. S., AND J. P. DUBEY. 1989. Immunohistochemical diagnosis of *Neospora caninum* in tissue sections. *American Journal of Veterinary Research* **50**: 1981–1983.
- LITERÁK, I., K. HEJLIČEK, J. NEZVAL, AND C. FOLK. 1992. Incidence of *Toxoplasma gondii* in a population of wild birds in the Czech republic. *Avian Pathology* **21**: 659–665.
- MCALLISTER, M. M., S. F. PARMLEY, L. M. WEISS, V. J. WELCH, AND A. M. MCGUIRE. 1996. An immunohistochemical method for detecting bradyzoite antigen (BAG 5) in *Toxoplasma gondii*-infected tissues cross-reacts with a *Neospora caninum* bradyzoite antigen. *Journal of Parasitology* **82**: 354–355.
- WALLACE, G. D. 1973. Intermediate and transport hosts in the natural history of *Toxoplasma gondii*. *American Journal of Tropical Medicine and Hygiene* **22**: 456–464.
- , L. MARSHALL, AND M. MARSHALL. 1972. Cats, rats and toxoplasmosis on a small Pacific island. *American Journal of Epidemiology* **95**: 475–482.
- WORK, T. M., J. G. MASSEY, B. A. RIDEOUT, C. H. GARDINER, D. B. LEDIG, O. C. H. KWOK, AND J. P. DUBEY. 2000. Fatal toxoplasmosis in free-ranging endangered 'Alala from Hawaii. *Journal of Wildlife Diseases* **36**: 205–212.

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Radical Curative Efficacy of Five-Day Regimen of Primaquine for Treatment of *Plasmodium vivax* Malaria in India

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ABSTRACT: For over 4 decades the antimalarial program in India has been prescribing a 5-day primaquine regimen as an antirelapse therapy to treat *Plasmodium vivax* malaria. In view of conflicting reports on the effectiveness of this regimen in the Indian subcontinent, and the varying prevalence of *P. vivax* in various ecosystems in India, the antirelapse efficacy of this regimen was evaluated in Orissa, a malaria endemic state in eastern India where *P. falciparum* predominates. In 723 cases of *P. vivax* infection treated with chloroquine alone and followed up weekly for 1 yr, the prevalence of recurrence of parasitaemia with fever was 8.6%. Among another 759 *P. vivax* cases treated with chloroquine and a 5-day regimen of primaquine at 15 mg/day (adult dose), the recurrence of infection was 6.5%. The difference in recurrence was not significant ($P = 0.53$). It is important to note that in a great majority of cases of *P. vivax* in this area, infection did not recur even without treatment with primaquine. This finding, that the use of the 5-day primaquine regimen with chloroquine had no significant advantage over the use of chloroquine alone, undermines the rationale of using primaquine as an antirelapse drug in forested areas with a high prevalence of *P. falciparum*.

Plasmodium vivax is the predominant human malarial species in most parts of India, except in the forested areas of central, eastern, and north-eastern India where *P. falciparum* is the main agent of infection. *Plas-*

modium vivax accounts for over two-thirds of the 3 million cases of malaria reported annually in India. For radical cure of *P. vivax* the malaria eradication program of India, started in 1958, adopted a regimen comprising 600 mg chloroquine given once and 15 mg primaquine for 5 consecutive days (adult doses). Contemporary Indian studies reported a low relapse rate in cases of *P. vivax* treated with chloroquine and a 5-day regimen of primaquine (Singh et al., 1954; Sharma et al., 1973). These studies, however, had the limitation of not comparing the efficacy of this regimen with a control group not treated with primaquine. Another study in Mysore State (now Karnataka State), however, demonstrated a low relapse rate with chloroquine plus 5-day primaquine therapy compared with chloroquine alone (Basavaraj, 1960). In contrast, through experimental infection of nonindigenous human volunteers with a West Pakistan strain of *P. vivax*, Contacos et al. (1973) found that the 5-day primaquine therapy was a 100% failure. In view of variations in the prevalence of *P. vivax* in different ecosystems in India and conflicting reports on the usefulness of the 5-day primaquine therapy in the Indian subcontinent, a study was conducted to evaluate the radical curative efficacy of the 5-day primaquine plus chloroquine therapy in treating *P. vivax* malaria in comparison with chloroquine alone.

The study was conducted in the villages of the Bisra block in a hill-forest ecosystem in Sundargarh district, Orissa State, in eastern India during 1988–1991. People live in small villages where the population